



Review

Molecular Imagine of Glutamate Receptor: Updated Statement

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1. Introduction:

Glutamic corrosive otherwise called glutamate previously recognized inside the bugs [1,2]. In the mammalian mind it contain approx. 10 μ M focus inside the mind [3], Glutamate most overflow and its existences the two spots inside cerebrum and alongside synapse pool [4]. Right around a few articulation of receptor for glutamate in glia and neurons other than heredities through assessment. Glutamate receptor articulation fall into two classification which is ionotropic ligands channels another is metabotropic particles channels. Its additionally partitioned into excitatory sorts I (mGluR1/mGluR5), another is type II receptors named Inhibitory nature types, and the sort III receptors. There are different excitatory person of Amino acids as found a few ID like NMDA(N-Aspartate D-methyl) receptors, Kainate, AMPA too additionally passable to cell layer. Considering this immensely assorted pharmacology, no specific or particular activity can be ascribed to glutamate receptors the sensory system. Synaptic excitatory administrative which is additionally evolved neuronal circuits which is more unmistakable activity on a few activities.

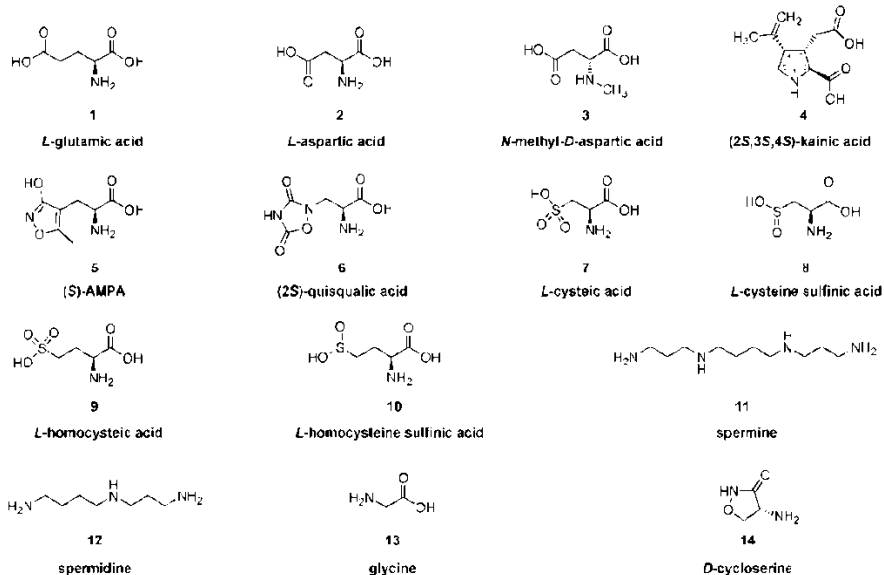


Figure 1. Chemical structures of excitatory amino acids (EAAs), sulfur-containing amino acids and polyamines.

A few survey on ionotropic receptor pharmacotherapeutical ways[5,6] and alongside metabotropic glutamate receptors [7,8]. There are a few continuous writing review accessible on Alzheimer and dementia alongside associated with its pathophysiology and fix of intriguing sicknesses, for example, [9], Parkinson's infection [10], ischemic stroke [11], and immune system illnesses of the sensory system [13], misery [17,18], and substance misuse/compulsion [19]. Its availability by its few focuses for little atoms imaging by single photon emanation registered tomography (SPECT) alongside positron discharge tomography (PET). While there have been a few surveys of this subject as of late [20,21,22,23].

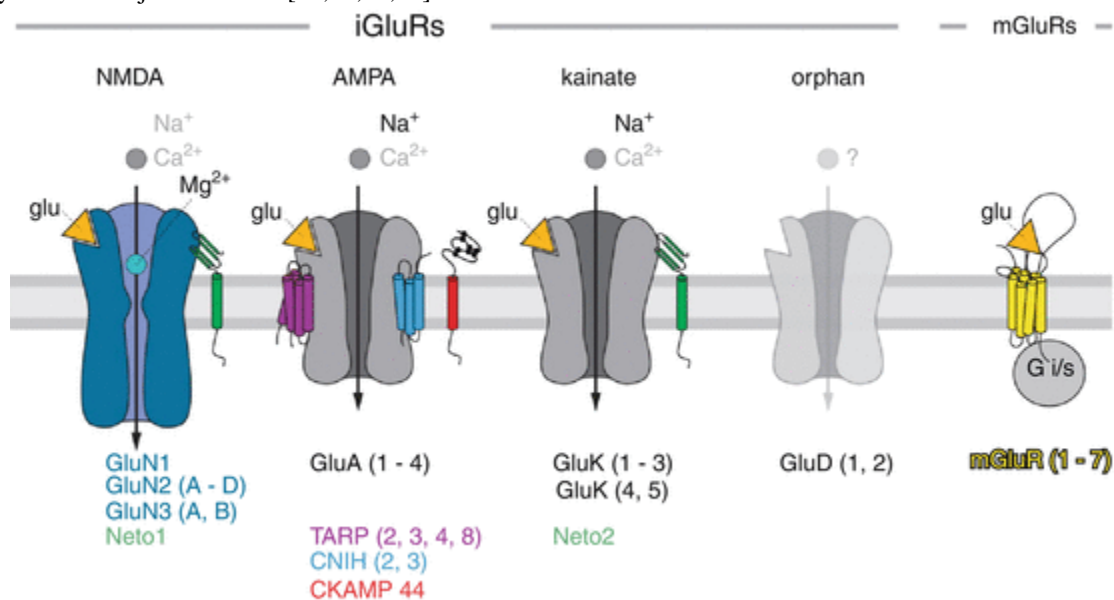


Fig. 2 Glutamate receptor Isomer.

A Concise Note on the Different Endpoints and Units of Restricting Examinations The correlation of discoveries with different PET and SPECT tracers in vivo requires a few thought of the different units and measures used to evaluate radiotracer take-up and restricting in the living mind. A neurochemist can quantify the limiting to layers or mind segments of the radio ligand over a scope of fixations in vitro. Subsequent to taking away the vague restricting,

the neurochemist can then work out the immersion restricting boundaries, to be specific the B_{max} , which is the outright convergence of the limiting locales in the example the proclivity or half-soaking ligand fixation. Here, the experimentalist has full control of the free ligand focuses and hatching conditions, which for the most part don't change during the limiting trial.

In PET and SPECT investigations of the living organic entity, the radiotracer disseminates all through the different tissues of the body after its organization as an intravenous bolus infusion. This promptly presents a period reliance of the fixation in blood, and further impacts of hepatic digestion and renal end on the radiotracer bioavailability. In the most straightforward type of quantitation, the PET or SPECT instrument reports the time-radioactivity bend in semiquantitative units of standard take-up esteem (SUV). This has units of level of the complete infused portion per gram of cerebrum tissue, at times with scaling to the comparing take-up in a reference tissue without any trace of explicit restricting (SUVR), on the off chance that such a tissue exists for a given objective. The SUV is a period subordinate boundary and is besides a composite record of explicit and vague restricting. In any case, SUV fills in as a helpful marker of how really the tracer can cross the blood-cerebrum hindrance (BBB). Fruitful SPECT or PET tracers as a rule achieve a SUV of something like 1% of the complete infused portion per gram of rat cerebrum and show higher SUV in mind districts enhanced with the designated restricting site. Knowing the blood vessel input capability by sequential testing with adjustment for radioactive metabolites, compartmental examination of the unique cerebrum bend gives evaluations of microparameters in mind. Boss among these are the unidirectional blood-mind freedom of the radiotracer (K_1), which has units of blood stream ($\text{ml g}^{-1} \text{min}^{-1}$), the fragmentary rate consistent for leeway of unbound tracer from cerebrum (k_2 ; min^{-1}), and the affiliation/separation rate constants with the cerebral objective restricting locales (k_3/k_4 ; min^{-1}). Compartmental examination additionally gives evaluations of macroparameters, which are the composite of a few microparameters, eminently the net blood-mind freedom comprehensive of irreversible catching in cerebrum (Family; $\text{mL g}^{-1} \text{min}^{-1}$), which is characterized as $(K_1 * k_3)/(k_2 + k_3)$.

Determined by straight graphical (Patlak-Gjedde) examination of dynamic PET/SPECT information, Family is very particular from the comparably named hindrance steady (K_i ; mols per liter) got from an impeding report in vitro, which is more likened to the IC_{50} , i.e., the plasma centralization of a contender dislodging half of the particular ligand restricting in cerebrum. Consistent state dissemination volume proportions of a tracer fixation in cerebrum to that in the blood (mL g^{-1}) incorporate the vague restricting (VND), which is equivalent to the proportion K_1/k_2 , and the all out circulation volume (VT), which likewise incorporates the particular restricting part, characterized by the proportion k_3/k_4 . The dimensionless restricting potential (BPND) addresses explicit restricting as $[(VT - VND)/VND]$. This profits us to the primary standards of autoradiography, in that the BPND got from PET estimations ought to be corresponding to the proportion of the immersion restricting boundaries B_{max}/KD estimated in vitro. On the off chance that there is a cerebrum district without any trace of explicit restricting, this can serve for the reference tissue computation of BPND, subsequently keeping away from the requirement for blood vessel inspecting.

2. Glutamate Receptors

2.1. Ionotropic Glutamate Receptors (iGluR)

2.1.1. Pharmacology of NMDA Receptor Ligands

The primary period of sub-atomic cerebrum imaging dates to the last part of the 1970s, with introductory endeavors to plan radiopharmaceuticals named with iodine-123 for SPECT, or with fleeting positron-emanating radionuclides carbon-11 ($t_{1/2}$ of 20 min) or fluorine-18 ($t_{1/2}$ of 109 min) for PET cerebrum imaging. The extraordinary greater part of synapse sub-atomic imaging reads up have utilized radioligands for dopamine, serotonin, and narcotic restricting locales, and progress in creating glutamate receptor ligands has gotten pace just lately. The main glutamate receptor studies designated the NMDA receptor, which is a tetraheteromer comprising of two mandatory GluN1 and two GluN2 as well as GluN3 subunits, framing together a transmembrane pore for $\text{Na}^+/\text{Ca}^{2+}$ deluge and K^+ efflux. The receptor subunits are encoded by seven qualities, with a solitary quality for GluN1, its record being dependent upon elective grafting. Four GluN2 subunits (GluN2A, 2B, 2C, 2D) and two GluN3 subunits (GluN3A, 3B) encoded by various qualities, individually, further add to receptor variety. There are many join variations and a few GluN2 records, however the NMDA receptors are for the most part liable to allosteric regulation by Zn^{2+} , the

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polyamines spermine (11) and spermidine (12), different proteins, and voltage subordinate barricade of the cation channel by Mg²⁺ (for survey, [24]). Actuation of the NMDA receptor requires restricting of the co-substrate amino corrosive L-glycine (13) to the GluN1 subunit and glutamate (1) (or NMDA, 3) to the GluN2 subunit, at explicit destinations in the extracellular area. Notwithstanding NMDA (3) and glutamate (1), different agonists of NMDA receptors incorporate D-cycloserine (14) (at the glycine-restricting site), homocysteic corrosive (9), the Amanita muscaria poison, ibotenic corrosive (15), and quinolinic corrosive (16). Among its main bad guys are the endogenous tryptophan metabolite, kynurenic corrosive (17), the over-the-counter hack suppressant, dextromethorphan (19), and the antihypertensive specialist if enprodil (21).

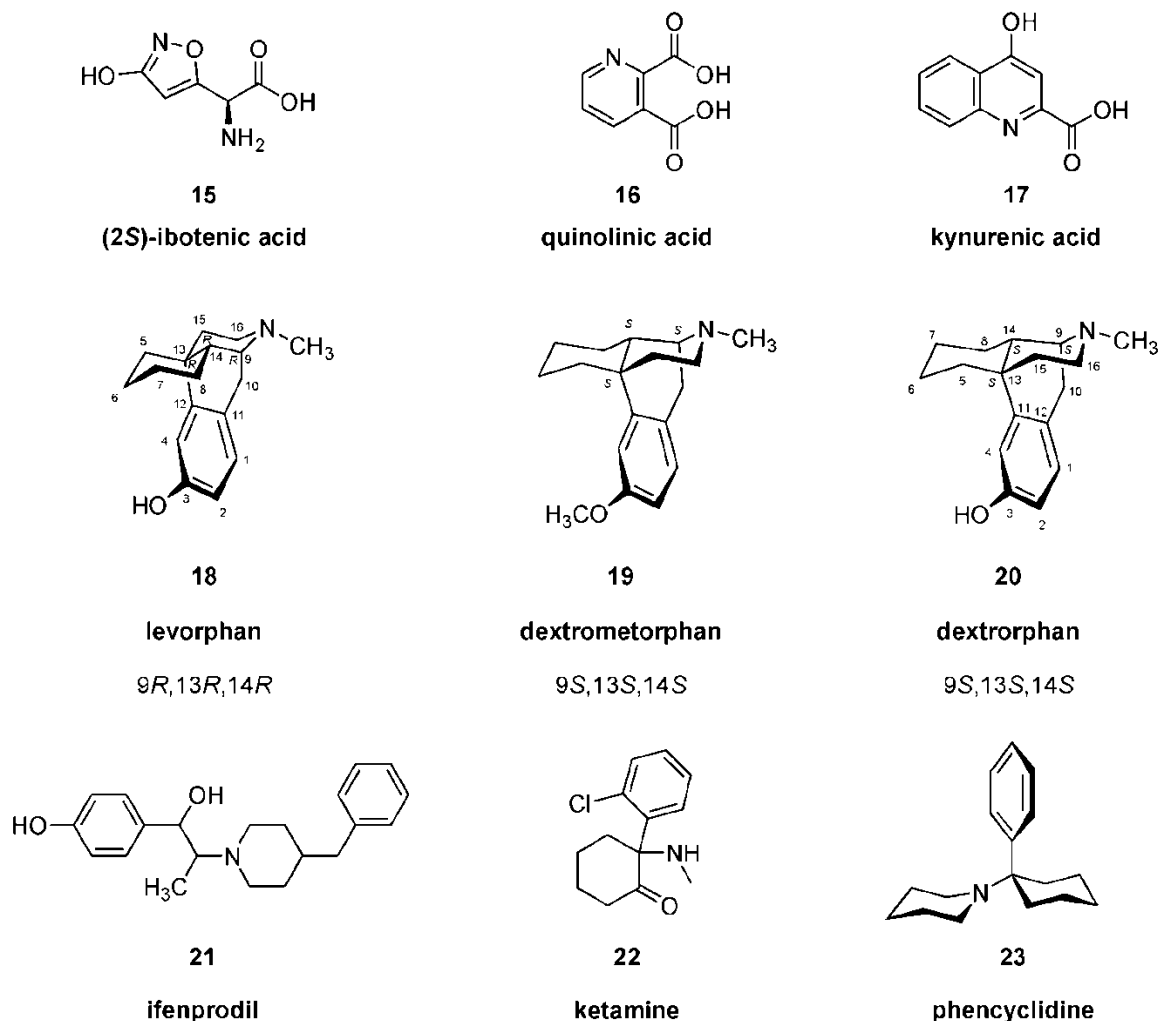


Figure:3 Chemical structures of some prototypic *N*-methyl-D-aspartate (NMDA) receptor ligands.

As a commonplace ligand gated particle channel, the NMDA receptor has an extracellular ligand-restricting space and a particle leading transmembrane area, which is just open upon receptor initiation. The NMDA receptor is pharmacologically mind boggling, introducing somewhere around four unmistakable restricting destinations, which are targetable by particular classes of ligands. These locales incorporate (A) the phencyclidine (PCP)/MK801 restricting site in the transmembrane pore, (B) the glutamate restricting destinations on the extracellular side GluN₂, (C) the co-agonist restricting site for glycine (13) on the GluN₁ subunit, and (D) allosteric modulator locales at the connection point of the dimers including the total tetramer. These destinations present possible focuses for PET/SPECT imaging. To this rundown could likewise be added the polyamine restricting site, which regulates the limiting of [3H]MK801 [29] and other intrachannel ligands.

While the manufactured narcotic subsidiaries levorphan (18, Hoffmann-La-Roche, 1952), dextrometorphan (19, Hoffmann-La-Roche, 1954), and dextrorphan (20) share the equivalent morphinan-skeleton, they vary fundamentally in their pharmacological profiles concerning selectivity for the PCP/MK801 restricting site in the NMDA channel. These mixtures were blended from cyclohexanone in a ten-step strategy [30]. Levorphan (18), which has a R-outright design by any means of its uneven focuses, is a powerful narcotic receptor agonist with a roughly seven-crease higher proclivity than morphine [31]. Dextrometorphan (19), which is the (9S,13S,14S)-isomer, has no pain relieving movement, yet is ordinarily utilized as hack suppressant. Dextromethorphan (19) and its primary metabolite dextrorphan (20) likewise show anticonvulsant and neuroprotective impacts, which are clearly gotten by enmity of NMDA glutamate receptors. In this way, stereochemistry of morphinan compounds decides their selectivity among narcotic and NMDA receptors, and, as introduced beneath, additionally as for sigma receptors.

The prototypic NMDA bad guy ligand [3H]MK801 ties to within the channel with a KD of 6 nM and Bmax of 250 nM in rodent cerebrum films [32]. Its limiting in cryostat areas from gerbil mind was most bountiful in the hippocampus and cerebral cortex, and was considerably uprooted by ketamine (22) and phencyclidine (23), however was unaffected by glutamate (1) or NMDA (3) [33]. The progress of [3H]MK801 as a ligand in vitro prompted the testing of [18F]fluoro-methyl-MK801 by PET (25), which showed somewhat heterogeneous take-up in cerebrum of living mandrill [34]. In any case, its limiting in vivo was not displaceable by challenge with phencyclidine (23) or overabundance non-radioactive MK801 (24), demonstrating an absence of explicit restricting.

2.1.2 NMDA Allosteric Modulators

N,N-dimethyl-2-(1*H*-pyrrolo[3,2-*b*]pyridin-1-yl)acetamide allosteric modulator ligands of the GluN2B subunit were prepared as potential PET tracers, among which [¹¹C]N2B-1810 showed “moderate” displaceable auto-radio graphic binding in rat telencephalon, and absent binding in cerebellum [32]. However, the tracer showed low (0.2% ID/g) and spatially uniform uptake in brain of living rats.

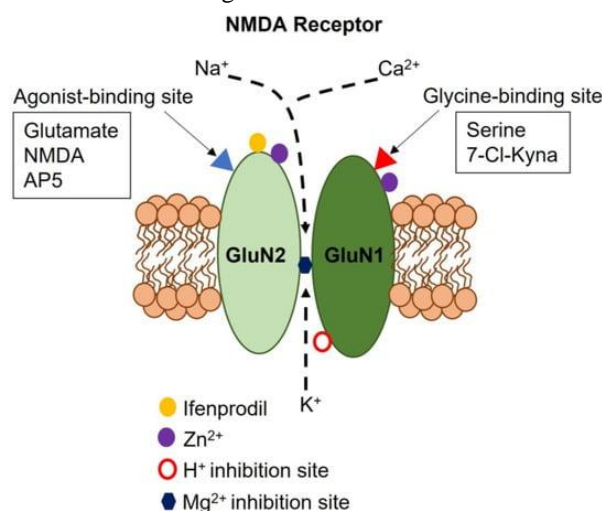


Fig. 4 The Emerging Role of N-Methyl-D-Aspartate (NMDA) Receptors in the Cardiovascular System:

2.1.3 AMPA Receptors

Previously known as quisqualate receptor, the AMPA receptor is so-named for its specific agonist, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic corrosive (5). For pertinent designs, see Figure 5. The AMPA receptor is a ligand gated particle channel made out of four kinds of subunits, assigned GluA1, 2, 3, and 4 (for survey, [33]). Like the NMDA receptor, AMPA receptors are dimers of dimers, made out of a GluA2 dimer and one of different dimers; changes in their synthesis, utilitarian properties and dealing across improvement basically decide synaptic versatility [34-45].

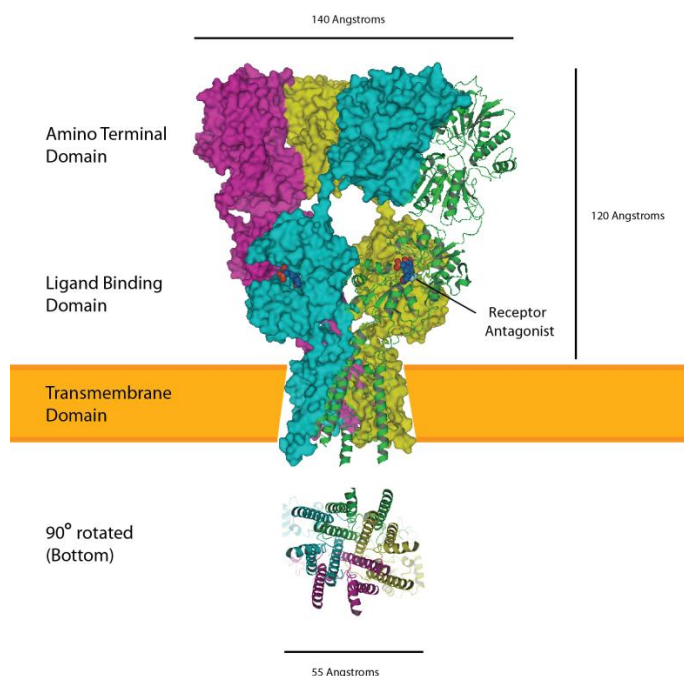


Figure: 5 NMDA Receptor that connects ion channels to deliver physiological action.

Dissimilar to NMDA receptors, every subunit of the AMPA receptor can tie its agonist, and pore opening expects restricting to at least two subunits. The GluA2 subunit grants to the channel a low porousness to Ca^{2+} , so the open channel just backings Na^{+}/K^{+} transition, and besides a voltage-subordinate restricting of polyamines to the GluA2 subunit balances the K^{+} current. AMPA actuation acts working together with NMDA receptors to intercede long haul synaptic potentiation, by which starting depolarization because of AMPA channel opening ousts the Mg^{2+} that would somehow impede NMDA receptor-intervened flows. This permits a flood of Ca^{2+} that eventually upregulates AMPA articulation to build the responsiveness of the film to glutamate.

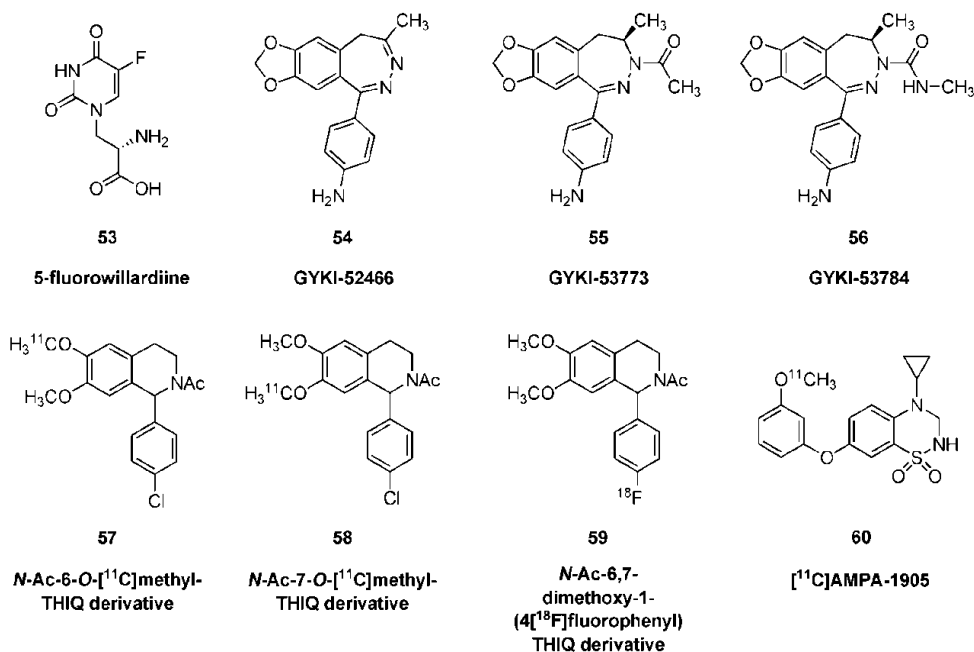


Figure 6: Structures of selected AMPA receptor ligands and non-competitive AMPA antagonists with 2,3-benzodiazepine scaffold.

2.1.4. Kainate Receptors

The kainate receptors are ionotropic non-NMDA receptors, which are selective to activation by the algal excitotoxic compound kainic acid. Kainate (4) is the most toxic of the excitatory amino acids, with a rank order of potency upon injection to the hippocampus kainate > ibotenate > NMDA > dihydrokainate > *D,L*-homocysteate > *L*-cysteate > *L*-aspartate > *L*-glutamate [46]. As such, kainic acid is a useful tool for neurochemical lesion studies. Like the other ionotropic glutamate receptors, the kainate receptor is a homomeric or heteromeric tetramer assembled from five possible subunits [47]. The ion channel of the kainate receptor is permissive to Na⁺/K⁺ flux, but the receptor seems also to possess coupling with a G-protein cascade, which is an unusual property for ionotropic receptors. Native kainate receptors expressed on neurons mediate slow excitation, distinct from the rapid depolarizations mediated by NMDA and AMPA receptors [48-56].

This property may be critically involved in encoding of temporal information and modulation of local and network spike activity, under the influence of auxiliary proteins like neuropilin and tolloid-like protein 1 (Neto1).

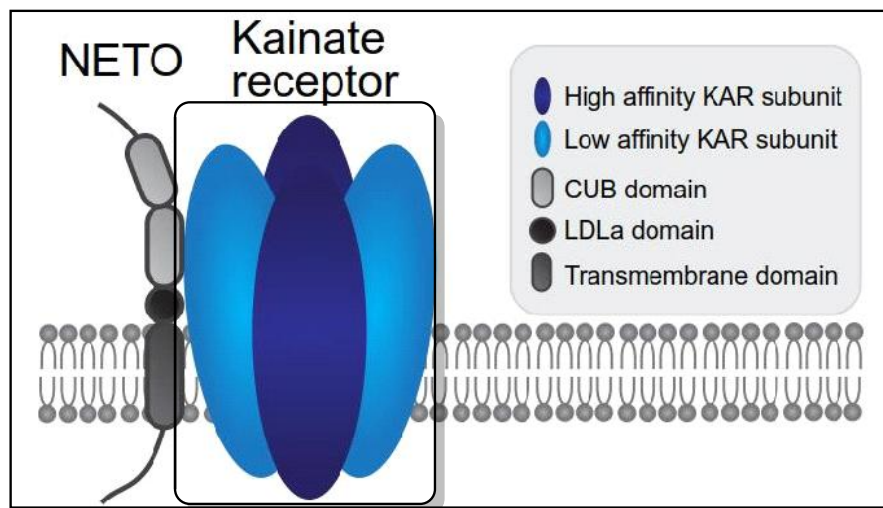


Fig.7 Kainate receptor and its physiological receptor.

3. Conclusions and Outlook

We have reviewed the present state of development of PET/SPECT probes for glutamate receptor imaging, including both ionotropic and metabotropic receptors, and discussed the suitability of the various tracers for reliable quantitation in the living brain. The diverse pharmacology of glutamate receptors, together with the central role of glutamatergic neurotransmission in brain function present a multitude of targets for molecular imaging. Quantitative imaging of ionotropic receptors is still difficult, despite early success with the intra-channel NMDA receptor SPECT ligand, CNS-1261 in clinical studies of schizophrenia. There has been some recent progress in developing suitable radioligands for PET imaging of the GluN2B subtype of the NMDA receptor.

There has been some success in preclinical development of ligands for AMPA receptors in the living brain, but molecular imaging of kainate receptors remains unattainable; this seems remarkable, given their central role in synaptic plasticity. There has been rather better progress in mGluR imaging, particularly for the mGluR5 subtype, which has a functional link to NMDA receptors via intracellular scaffolding proteins. This has enabled rather extensive clinical investigations of mGluR5 availability in disorders such as drug abuse/addiction, depression, and PTSD.

It is important to note that these disorders are heterogeneous and have high comorbidity with each other. For example, smoking history must be strictly controlled in PET studies with the most widely used mGluR5 ligand, [¹¹C] ABP688, and there is some evidence of rapid diurnal changes in mGluR5 availability. However, several members of the mGluR family including mGluR3, mGluR6, mGluR7, and mGluR8 remain uninvestigated by molecular imaging.

References:

1. Kerkut G.A., Walker R.J. The effect of *L*-glutamate, acetylcholine and gamma-aminobutyric acid on the miniature end-plate potentials and contractures of the coxal muscles of the cockroach, *Periplaneta Americana*. *Comp. Biochem. Physiol.* 1966;17:435–454.
2. Gerschenfeld H.M. Chemical transmission in invertebrate central nervous systems and neuromuscular junctions. *Physiol. Rev.* 1973;53:1–119.
3. Schousboe A. Transport and metabolism of glutamate and GABA in neurons and glial cells. In: Smythies J.R., Bradley R.J., editors. *International Review of Neurobiolog.* Volume 22. Academic Press; Cambridge, MA, USA: 1981. pp. 1–45.
4. Zhou Y., Danbolt N.C. Glutamate as a neurotransmitter in the healthy brain. *J. Neural. Transm. (Vienna)* 2014;121:799–817.
5. Traynelis S.F., Wollmuth L.P., McBain C.J., Menniti F.S., Vance K.M., Ogden K.K., Hansen K.B., Yuan H., Myers S.J., Dingledine R. Glutamate receptor ion channels: Structure, regulation, and function. *Pharmacol. Rev.* 2010;62:405–496.
6. Liu W., Jiang X., Zu Y., Yang Y., Liu Y., Sun X., Xu Z., Ding H., Zhao Q. A comprehensive description of GluN2B-selective *N*-methyl-*D*-aspartate (NMDA) receptor antagonists. *Eur. J. Med. Chem.* 2020;200:112447.
7. Niswender C.M., Conn P.J. Metabotropic glutamate receptors: Physiology, pharmacology, and disease. *Ann. Rev. Pharmacol. Toxicol.* 2010;50:295–322.
8. Roth B.L. Molecular pharmacology of metabotropic receptors targeted by neuropsychiatric drugs. *Nat. Struct. Mol. Biol.* 2019;26:535–544.
9. Wang R., Reddy P.H. Role of glutamate and NMDA receptors in Alzheimer’s disease. *J Alzheimers Dis.* 2017;57:1041–1048.
10. Zhang Z., Zhang S., Fu P., Zhang Z., Lin K., Ko J.K.-S., Yung K.K.-L. Roles of glutamate receptors in Parkinson’s disease. *Int. J. Mol. Sci.* 2019;20:4391.
11. Fern R., Matute C. Glutamate receptors and white matter stroke. *Neurosci. Lett.* 2019;694:86–92.
12. Hanada T. Iontropic glutamate receptors in epilepsy: A review focusing on AMPA and NMDA receptors. *Biomolecules.* 2020;10:464.
13. Levite M. Glutamate receptor antibodies in neurological diseases: Anti-AMPA-GluR3 antibodies, anti-NMDA-NR1 antibodies, anti-NMDA-NR2A/B antibodies, anti-mGluR1 antibodies or anti-mGluR5 antibodies are present in subpopulations of patients with either: Epilepsy, encephalitis, cerebellar ataxia, systemic lupus erythematosus (SLE) and neuropsychiatric SLE, Sjogren’s syndrome, schizophrenia, mania or stroke. These autoimmune anti-glutamate receptor antibodies can bind neurons in few brain regions, activate glutamate receptors, decrease glutamate receptor’s expression, impair glutamate-induced signaling and function, activate blood brain barrier endothelial cells, kill neurons, damage the brain, induce behavioral/psychiatric/cognitive abnormalities and Ataxia in animal models, and can be removed or silenced in some patients by immunotherapy. *J. Neural Transm. (Vienna)* 2014;121:1029–1075.
14. Balu D.T. The NMDA receptor and schizophrenia: From pathophysiology to treatment. In: Schwarcz R., editor. *Advances in Pharmacology.* Volume 76. Academic Press; Cambridge, MA, USA: 2016. pp. 351–382.
15. Dean B., Gibbons A.S., Boer S., Uezato A., Meador-Woodruff J., Scarr E., McCullumsmith R.E. Changes in cortical *N*-methyl-*D*-aspartate receptors and post-synaptic density protein 95 in schizophrenia, mood disorders and suicide. *Aust. N. Z. J. Psychiatry.* 2016;50:275–283.
16. Maksymetz J., Moran S.P., Conn P.J. Targeting metabotropic glutamate receptors for novel treatments of schizophrenia. *Mol. Brain.* 2017;10:15.
17. Murrough J., Abdallah C., Mathew S. Targeting glutamate signalling in depression: Progress and prospects. *Nat. Rev. Drug Discov.* 2017;16:472–486.

18. Kalmoe M.C., Janski A.M., Zorumski C.F., Nagele P., Palanca B.J., Conway C.R. Ketamine and nitrous oxide: The evolution of NMDA receptor antagonists as antidepressant agents. *J. Neurol. Sci.* 2020;412:116778.
19. Ueno F., Suzuki T., Nakajima S., Matsushita S., Mimura M., Miyazaki T., Takahashi T., Uchida H. Alteration in AMPA receptor subunit expression and receptor binding among patients with addictive disorders: A systematic review of human postmortem studies. *Neuropsychopharmacol. Rep.* 2019;39:148–155.
20. Mu L., Schubiger P.A., Ametamey S.M. Radioligands for the PET imaging of metabotropic glutamate receptor subtype 5 (mGluR5) *Curr. Top. Med. Chem.* 2010;10:1558–1568.
21. Majo V.J., Prabhakaran J., Mann J.J., Kumar J.S. PET and SPECT tracers for glutamate receptors. *Drug Discov. Today.* 2013;18:173–184.
22. Gruber S., Ametamey S.M. Imaging the glutamate receptor subtypes-Much achieved, and still much to do. *Drug Discov. Today Technol.* 2017;25:27–36.
23. Fu H., Chen Z., Josephson L., Li Z., Liang S.H. Positron Emission Tomography (PET) Ligand Development for Ionotropic Glutamate Receptors: Challenges and Opportunities for Radiotracer Targeting *N*-Methyl-*D*-aspartate (NMDA), α -Amino-3-hydroxy-5-methyl-4-isoxazolepropionic Acid (AMPA), and Kainate Receptors. *J. Med. Chem.* 2018;62:403–419.
24. Glasgow N.G., Siegler Retchless B., Johnson J.W. Molecular bases of NMDA receptor subtype-dependent properties. *J Physiol.* 2015;593:83–95.
25. Wyllie D.J.A., Livesey M.R., Hardingham G.E. Influence of GluN2 subunit identity on NMDA receptor function. *Neuropharmacology.* 2013;74:4–17.
26. Vieira M., Yong X.L.H., Roche K.W., Anggono V. Regulation of NMDA glutamate receptor functions by the GluN2 subunits. *J. Neurochem.* 2020;154:121–143.
27. Sun Y., Xu Y., Cheng X., Chen X., Xie Y., Zhang L., Wang L., Hu J., Gao Z. The differences between GluN2A and GluN2B signaling in the brain. *J. Neuro. Res.* 2018;96:1430–1443.
28. Sapkota K., Mao Z., Synowicki P., Lieber D., Liu M., Ikezu T., Gautam V., Monaghan D.T. GluN2D *N*-Methyl-*D*-aspartate receptor subunit contribution to the stimulation of brain activity and gamma oscillations by ketamine: Implications for schizophrenia. *J. Pharm. Exp. Ther.* 2016;356:702–711.
29. Williams K., Pullan L.M., Romano C., Powel R.J., Salama A.I., Molinoff P.B. An antagonist/partial agonist at the polyamine recognition site of the *N*-methyl-*D*-aspartate receptor that alters the properties of the glutamate recognition site. *J. Pharm. Exp. Ther.* 1992;262:539–544.
30. Casy A.F., Parfitt R.T. *Opioid Analgesics: Chemistry and Receptors.* Plenum Press; New York, NY, USA: 1986.
31. Mohacsi E., Leimgruber W., Baruth H. Synthesis and pharmacology of metabolically stable *tert*-butyl ethers of morphine and levorphanol. *J. Med. Chem.* 1982;25:1264–1266.
32. Wong E.H.F., Knight A.R., Woodruff G.N. [³H]MK-801 Labels a Site on the *N*-Methyl-*D*-Aspartate Receptor Channel Complex in Rat Brain Membranes. *J. Neurochem.* 1988;50:274–281.
33. Bowery N., Wong E., Hudson A. Quantitative autoradiography of [³H]-MK-801 binding sites in mammalian brain. *Br. J. Pharmacol.* 1988;93:944–954.
34. Blin J., Denis A., Yamaguchi T., Crouzel C., MacKenzie E.T., Baron J.C. PET studies of [¹⁸F]methyl-MK-801, a potential NMDA receptor complex radioligand. *Neurosci. Lett.* 1991;121:183–186.
35. Maragos W.F., Penney J.B., Young A.B. Anatomic correlation of NMDA and ³H-TCP-labeled receptors in rat brain. *J. Neurosci.* 1988;2:493–501.
36. Ferrarese C., Guidotti A., Costa E., Miletich R.S., Rice K.C., de Costa B.R., Fulham M.J., Di Chiro G. In vivo study of NMDA-sensitive glutamate receptor by fluorothienylcyclohexylpiperidine, a possible ligand for positron emission tomography. *Neuropharmacology.* 1991;30:899–905.
37. Ouyang X., Mukherjee J., Yang Z.-Y. Synthesis, radiosynthesis, and biological evaluation of fluorinated thienylcyclohexyl piperidine derivatives as potential radiotracers for the NMDA receptor-linked calcium ionophore. *Nucl. Med. Biol.* 1996;23:315–324.

38. Roger G., Lagnel B., Besret L., Bramoullé Y., Coulon C., Ottaviani M., Kassiou M., Bottlaender M., Valette H., Dollé F. Synthesis, radiosynthesis and In vivo evaluation of 5-[3-(4-Benzylpiperidin-1-yl)prop-1-ynyl]-1,3-dihydrobenzimidazol-2-[¹¹C]one, as a potent NR1A/2B subtype selective NMDA PET radiotracer. *Bioorg. Med. Chem.* 2003;11:5401–5408.
39. Shiue C.-Y., Vallabhahosula S., Wolf A.P., Dewey S.L., Fowler J.S., Schlyer D.J., Arnett C.A., Zhou Y.-G. Carbon-11 labelled ketamine-synthesis, distribution in mice and PET studies in baboons. *Nucl. Med. Biol.* 1997;24:145–150.
40. Hartvig P., Valtysson J., Antoni G., Westerberg G., Långström B., Moberg E.R., Øye I. Brain kinetics of (R)- and (S)-[N-methyl-¹¹C]ketamine in the rhesus monkey studied by positron emission tomography (PET) *Nucl. Med. Biol.* 1994;21:927–934.
41. Salabert A.-S., Fonta C., Fontan C., Adel D., Alonso M., Pestourie C., Belhadij-Tahar H., Tafani M., Payoux P. Radiolabeling of [¹⁸F]fluoroethylnormemantine and initial in vivo evaluation of this innovative PET tracer for imaging the PCP sites of NMDA receptors. *Nucl. Med. Biol.* 2015;42:643–653.
42. Salabert A.-S., Mora-Ramirez E., Beaurain M., Alonso M., Fontan C., Tahar H.B., Boizeau M.L., Tafani M., Bardiès M., Payoux P. Evaluation of [¹⁸F]FNM biodistribution and dosimetry based on whole-body PET imaging of rats. *Nucl. Med. Biol.* 2018;59:1–8.
43. Ametamey S.M., Samnick S., Leenders K.L., Vontobel P., Quack G., Parsons C.G., Schubiger P.A. Fluorine-18 radiolabelling, biodistribution studies and preliminary PET evaluation of a new memantine derivative for imaging the NMDA receptor. *J. Recept. Signal Transduct. Res.* 1999;19:129–141.
44. Ametamey S.M., Bruehlmeier M., Kneifel S., Kokic M., Honer M., Arigoni M., Buck A., Burger C., Samnick S., Quack G., et al. PET studies of ¹⁸F-memantine in healthy volunteers. *Nucl. Med. Biol.* 2002;29:227–231.
45. Dumont F., Sultana A., Waterhouse R.N. Synthesis and in vitro evaluation of N,N'-diphenyl and N-naphthyl-N'-phenylguanidines as N-methyl-D-aspartate receptor ion-channel ligands. *Bioorg. Med. Chem. Lett.* 2002;12:1583–1586.
46. Naumiec G.R., Jenko K.J., Zoghbi S.S., Innis R.B., Cai L., Pike V.W. N'-3-(Trifluoromethyl)phenyl Derivatives of N-Aryl-N'-methylguanidines as Prospective PET Radioligands for the Open Channel of the N-Methyl-D-aspartate (NMDA) Receptor: Synthesis and Structure-Affinity Relationships. *J. Med. Chem.* 2015;58:9722–9730.
47. Waterhouse R.N., Slifstein M., Dumont F., Zhao J., Chang R.C., Sudo Y., Sultana A., Balter A., Laruelle M. In vivo evaluation of [¹¹C]N-(2-chloro-5-thiomethylphenyl)-N'- (3-methoxy-phenyl)-N'-methylguanidine ([¹¹C]GMOM) as a potential PET radiotracer for the PCP/NMDA receptor. *Nucl. Med. Biol.* 2004;31:939–948. doi: 10.1016/j.nucmedbio.2004.03.012.
48. Klein P.J., Schuit R.C., Metaxas A., Christiaans J.A.M., Kooijman E., Lammertsma A.A., van Berckel B.N.M., Windhorst A.D. Synthesis, radiolabeling and preclinical evaluation of a [¹¹C]GMOM derivative as PET radiotracer for the ion channel of the N-methyl-D-aspartate receptor. *Nucl. Med. Biol.* 2017;51:25–32.
49. Haider A., Herde A.M., Krämer S.D., Varisco J., Keller C., Frauenknecht K., Auberson Y.P., Temme L., Robaa D., Sippl W., et al. Preclinical Evaluation of Benzazepine-Based PET Radioligands (R)- and (S)-¹¹C-Me-NB1 Reveals Distinct Enantiomeric Binding Patterns and a Tightrope Walk Between GluN2B- and σ 1-Receptor-Targeted PET Imaging. *J. Nucl. Med.* 2019;60:1167–1173.
50. Haider A., Iten I., Ahmed H., Müller Herder A., Gruber S., Krämer S.D., Keller C., Schibli R., Wunsch B., Mu L., et al. Identification and preclinical evaluation of a radiofluorinated benzazepine derivative for imaging the GluN2B subunit of the ionotropic NMDA receptor. *J. Nucl. Med.* 2018
51. Ahmed H., Haider A., Varisco J., Stanković M., Wallimann R., Gruber S., Iten I., Häne S., Müller Herde A., Keller C., et al. Structure-affinity relationships of 2,3,4,5-tetrahydro-1H-3-benzazepine and 6,7,8,9-tetrahydro-5H-benzo[7]annulen-7-amine analogues and the discovery of a radiofluorinated 2,3,4,5-tetrahydro-1H-3-benzazepine congener for imaging GluN2B subunit-containing N-methyl-D-aspartate receptors. *J. Med. Chem.* 2019;62:9450–9470.

52. Ahmed H., Wallimann R., Haider A., Hosseini V., Gruber S., Robledo M., Nguyen T., Herde A.M., Iten I., Keller C., et al. Preclinical development of ^{18}F -OF-NB1 for imaging GluN2B-containing *N*-methyl-*D*-aspartate receptors and its utility as a biomarker for amyotrophic lateral sclerosis. *J. Nucl. Med.* 2020
53. Schwartz D.E., Bruderer H., Rieder J., Brossi A. Metabolic studies of Versidyne, a new analgesic, in the rabbit and in man. *Biochem. Pharmacol.* 1964;13:777–778.
54. Jakobsson J.E., Gourni E., Khanapur S., Brito B., Riss P.J. Synthesis and characterization in rodent brain of the subtype-selective NR2B NMDA receptor ligand [^{11}C]Ro04-5595 as a potential radiotracer for positron emission tomography. *ACS Omega.* 2019;4:9925–9931.
55. Wilson A.A., Garcia A., Jin L., Houle S. Radiotracer synthesis from [^{11}C]iodomethane: A remarkably simple captive solvent method. *Nucl. Med. Biol.* 2000;27:529–532.
56. Cai L., Liow J.-S., Morse C., Telu S., Davies R., Frankland M., Zoghbi S., Cheng K., Hall M., Innis R.B., et al. Evaluation of ^{11}C -NR2B-SMe and its enantiomers as PET radioligands for imaging the NR2B subunit within the NMDA receptor complex in rats. *J. Nucl. Med.* 2020;61:1212–1220.
